## Mycobacterium Tuberculosis-Specific Antigens As Subunit Vaccines Against Tuberculosis

## Mustafa AS

Kuwait University, Kuwait

Tuberculosis (TB) is a chronic infectious disease caused by the bacterium Mycobacterium tuberculosis. TB is present globally and ranks among the top 10 causes of all deaths and it is the leading cause of death from a single infectious agent, with a ranking above HIV/AIDS [1]. According to the worldwide estimates from the World Health Organization (WHO), about 10 million people became ill with TB and 1.5 million people died of it in 2018 [1]. The situation of TB is further complicated as a public health problem due to drug-resistance and increased susceptibility of HIV/AIDS patients. In 2018, there were about 0.5 million new cases of TB resistant to rifampicin (the most effective first-line anti-TB drug) and 0.4 million of them had multidrug resistant TB Moreover, about 25% of the global population is estimated to be latently infected with M. tuberculosis, and 5 - 15% of them have the lifetime risk of developing clinically active TB disease. An efficient and effective TB control program requires development of effective drugs against resistance strains, cost-effective technologies for the specific diagnosis of latent and active disease and effective, safe and affordable vaccines. Among the various possibilities, the availability of new TB vaccines deserves priority because cost-effective and safer vaccines are the best option as such vaccines have been effective in controlling and eradicating many infectious diseases.

The currently available vaccine against TB is a live attenuated strain of pathogenic Mycobacterium bovis known as Bacillus Calmette-Guérin (BCG). Although BCG has been extensively used to vaccinate against TB since 1921 and it is the most widely used vaccine in the world, it is the most controversial vaccine in current use. This is because BCG has failed to consistently protect against the major manifestation of TB in adults, i.e. pulmonary TB [4]. The variations in protection have ranged from nil (e.g. in India and Malawi) to 80% (e.g. in the United Kingdom). Furthermore, as BCG is a live mycobacterium, it is not suitable for

vaccinating immuno compromised individuals, particularly patients suffering from HIV/AIDS, due to the possibility of causing disease in such individuals. Since BCG vaccination induces positivity to the commonly used tuberculin skin test for the diagnosis of TB, it becomes difficult to use this test for diagnostic epidemiological investigations or in populations vaccinated with BCG. In addition, antigens cross-reactive BCG has with mycobacteria environmental leading to masking or blocking effects. According to masking hypothesis, early sensitization with environmental mycobacteria confers some level of protection against TB that masks the effect of a vaccine given later in life due to presence of cross-reactive antigens. Whereas, the blocking hypothesis postulates that previous immune response to cross-reactive antigens, because of environmental mycobacteria sensitization, prevents vaccine taking of a new TB vaccine [5]. The use of M. tuberculosis-specific antigens may overcome these effects. Therefore, to have vaccines better than BCG, development of new subunit vaccines based on M. tuberculosis-specific antigens is needed.

The identification of *M. tuberculosis*-specific antigens has been facilitated by advances in mycobacterial genome sequencing and the genomics to identify comparative М. tuberculosis-specific genomic regions. Such studies have identified 11 M. tuberculosisspecific genomic regions known as regions of differences (RDs), which are deleted/absent in all BCG substrains currently used in different parts of the world to vaccinate against TB [7]. In silico analysis has suggested that these RDs can potentially encode 89 proteins. To identify the candidate proteins suitable for vaccine development, it is essential to identify the immunodominant proteins from the RDs that can mediate protection against TB.

Protection in TB is primarily mediated by cellular immunity involving the interaction of antigen-specific T cells and macrophages. This interaction is often indicated by antigeninduced proliferation of T cells and is

2

dependent on the interplay of cytokines secreted by these cells. Although a broad spectrum of cytokines contribute to protection, the T-helper type 1 (Th1) cytokines, dominated by secretion of IL-2 (responsible for proliferation of antigen-reactive T cells) and (responsible IFN-v for activation of macrophages to kill ingested bacilli), are considered principal mediators of protective immunity against TB. Therefore, Th1 cell reactivity, indicated by antigen-induced cell proliferation and secretion of IFN-y, has been used as a marker to detect antigens involved in protective immunity and thus suitable for the identification of candidate vaccines.

The screening of *M. tuberculosis*-specific antigens with human peripheral blood cells and naturally infected cattle identified a total of six antigens inducing strong Th1-responses, i.e. PE35, PPE68, ESAT-6, CFP10, Rv3619c and Rv2346c. Testing of these antigens in animal models of TB have shown that all of these antigens are capable of inducing Th1 responses when used along with appropriate adjuvants and delivery systems like DNA non-pathogenic vaccine vectors and mycobacteria, including BCG. In mice and guinea pigs, immunizations with preparations containing ESAT-6, CFP-10 and Rv3619c have provided protection against challenges with virulent *M. tuberculosis*.

In humans, a recombinant subunit tuberculosis vaccine candidate containing dextran-binding domain modified Ag85a and ESAT6-CFP10 M. tuberculosis antigens with an adjuvant has been tested in phase I clinical trial. Safety and immunogenicity of this vaccine were determined in an open-label clinical trial on healthy volunteers previously vaccinated with BCG. The candidate vaccine had an acceptable safety profile and was welltolerated. Furthermore, the immunization protocols lead to a significant increase in the concentration of the Th1-cytokine IFN-y and IgG antibodies. The immune responses were induced to all antigens included in the vaccine preparation.

In conclusion, several *M. tuberculosis*-specific antigens induce protective Th1-type immune responses with human cells *in vitro*. The immunogenicity and protective efficacy studies in animals have shown that these antigens may be useful as subunit vaccine candidates against TB. In a human phase I trial, a preparation containing *M. tuberculosis*-specific antigens has been found immunogenic to induce both cellular and humoral immunity. Further studies are required to determine the protective efficacy in phase II and Phase III clinical trials in humans to determine the protective efficacy of the candidate vaccine.

## References

- 1. World Health Organization. Tuberculosis. <u>https://www.who.int/news-room/fact-</u> <u>sheets/detail/tuberculosis</u> (Accessed on August 6, 2020)
- Smith R: Eradication of tuberculosis by 2050 impossible without new vaccine. BMJ 2009;338:b1291.
- <u>Toumi</u> M, <u>Ricciardi</u> W. The Economic value of vaccination: Why prevention is wealth. <u>J Mark Access Health</u> Policy. 2015; 3: 10.
- 4. Mustafa AS. BCG pros and cons and new/improved vaccines for tuberculosis. Text Book of Biochemistry, Biotechnology, Allied Molecular Medicine, Fourth and Edition. Editors: Talwar GP, Hasnain SE, Sarin SK & Sayed Hasnain, PHI Learning Private Publisher: Limited, Delhi, India. Chapter 117, 2016, pp 1347-1353.
- Arregui S, Sanz J, Marinova D, Martín C, Moreno Y. On the impact of masking and blocking hypotheses for measuring the efficacy of new tuberculosis vaccines. *Peer J*. 2016;4:e1513].
- Mustafa AS, Al-Attiyah R. Tuberculosis: Looking beyond BCG vaccines. J Postgrad Med 2003; 49:129-140.
- Mustafa AS. Mycobacterial gene cloning and expression, comparative genomics, bioinformatics and proteomics in relation to the development of new vaccines and diagnostic reagents. Med Princ Pract 2005; 14(suppl 1): 27–34.
- 8. Mustafa AS. Development of new vaccines and diagnostic reagents against tuberculosis. Mol Immunol 2002; 39:113-119.
- 9. Mustafa AS. What's new in the development of tuberculosis vaccines. Med Princ Pract 2012; 21:195-196.
- 10. Mustafa AS. T-helper 1, T-helper 2, pro-inflammatory and anti-

inflammatory cytokines in tuberculosis. IJMPBS 2014; 3: 1-14

- 11. Mustafa AS. Immune responses to candidate vaccine antigens delivered through naked plasmid and mycobacterial vectors. Open Conf Proc J 2016; 7:153-199.
- 12. Mustafa AS. Biotechnology in the development of new vaccines and diagnostic reagents against tuberculosis. Curr Pharm Biotechnol 2001; 2: 157-173.
- 13. Mustafa AS. Vaccine potential of *Mycobacterium tuberculosis*-specific genomic regions: *in vitro* studies in humans. Expert Reviews Vaccines 2009; 8:1309-12.
- 14. Mustafa AS. Antigens for DNA vaccines against tuberculosis. Mycobacterial Dis 2018; 8:1.
- 15. Mustafa AS. Cell mediated immunity assays identify proteins of diagnostic and vaccine potential from genomic regions of difference of *Mycobacterium tuberculosis*. Kuwait Med J 2010; 42:98-105.
- Mustafa AS. Diagnostic and vaccine potentials of ESAT-6 family proteins encoded by *M. tuberculosis* genomic regions absent in *M. bovis* BCG. J Mycobac Dis 2013; 3:129.
- 17. Mustafa AS, Cockle PJ, Shaban F, Hewinson RG, Vordermeier HM. Immunogenicity of RD1 region gene products in *M. bovis* infected cattle. Clin Exp Immunol 2002; 130:37-42.
- Mustafa AS, Shaban FA, Al-Attiyah RJ, Abal AT, El-Shamy AM, Andersen P, Oftung F (2003). Human Th1 cell lines recognize the Mycobacterium tuberculosis ESAT-6 antigen and its peptides in association with frequently expressed HLA class II molecules. Scand J Immunol 2003; 57:125-134.
- 19. Mustafa AS. Recombinant and synthetic peptides to identify *Mycobacterium tuberculosis* antigens and epitopes of diagnostic and vaccine relevance. Tuberculosis (Edinb) 2005; 85:367-76.
- 20. Mustafa AS, Shaban FA. ProPred analysis and experimental evaluation of promiscuous Th1 cell epitopes of three major secreted antigens of *Mycobacterium tuberculosis*. Tuberculosis (Edinb) 2006; 86:115-24.
- 21. Al-Attiyah R, Mustafa AS. Characterization of human cellular

immune responses to novel *Mycobacterium tuberculosis* antigens encoded by genomic regions absent in *Mycobacterium bovis* BCG. Infect Immun 2008; 76:4190-4198.

Mustafa AS.

- 22. Mustafa AS, El-Shamy AM, Madi NM, Amoudy HA, Al-Attiyah R. Cell mediated immune responses to complex and single mycobacterial antigens in tuberculosis patients with diabetes. Med Princ Pract 2008; 17:325-330.
- Mustafa AS, R Al-Attiyah, SNM Hanif, FA Shaban. Efficient testing of pools of large numbers of peptides covering 12 open reading frames of *M. tuberculosis* RD1 and identification of major antigens and immunodominant peptides recognized by humanTh1 cells. Clin Vaccine Immunol 2008; 15:916-924.
- 24. Hanif SNM, El-Shammy AM, Al-Attiyah R, Mustafa AS. Whole blood assays to identify Th1 cell antigens and peptides encoded by *Mycobacterium tuberculosis*-specific RD1 genes. Med Princ 2008; Pract17:244-249.
- 25. Mustafa AS, Al-Saidi F, El-Shamy ASM and Al-Attiyah R. Cytokines in response to proteins predicted in genomic regions of difference of *Mycobacterium tuberculosis*. Microbiol Immunol 2011; 55:267-278.
- 26. Mustafa AS. In silico analysis and experimental validation of Mycobacterium tuberculosis-specific proteins and peptides of Mycobacterium tuberculosis for immunological diagnosis and vaccine development. Med Princ Pract 2013; 22 (Suppl 1): 43-51.
- 27. Mustafa AS. Characterization of a cross-reactive, immunodominant and HLA-promiscuous epitope of *Mycobacterium tuberculosis*-specific major antigenic protein PPE68. PLoS One 2014; 9:e103679.
- 28. Hanif SNM, Al-Attiyah R and Mustafa AS. DNA vaccine constructs expressing *Mycobacterium tuberculosis*-specific genes induce immune responses. Scand J Immunol 2010; 72:408-415.
- 29. Hanif SN, Al-Attiyah R, Mustafa AS. Cellular immune responses in mice induced by *M. tuberculosis* PE35-DNA vaccine construct. Scand J Immunol 2011; 74:554-60.

4

- Shaban K, Amoudy HA, Mustafa AS. Cellular immune responses to recombinant *Mycobacterium bovis* BCG constructs expressing major antigens of region of difference 1 of *Mycobacterium tuberculosis*. Clin Vaccine Immunol 2013; 20:1230-1237.
- 31. Amoudy HA, Ebrahimi BH, Mustafa AS. Immune responses against *Mycobacterium tuberculosis*-specific proteins PE35 and CFP10 in mice immunized with recombinant

*Mycobacterium vaccae*. Saudi Med J 2014; 35:350-359.

- 32. Safar HA, Mustafa AS, Amoudy HA, El-Hashim A. The effect of adjuvants and delivery systems on Th1, Th2, Th17 and Treg cytokine responses in mice immunized with Mycobacterium tuberculosis-specific proteins. PLoS One 2020;15:e0228381.
- 33. Mustafa AS. Vaccine potential of mycobacterial antigens against asthma. Med Princ Pract.